

### **Review**

## The epigenetic impact of the exposome on Sjögren's disease. A systematic review

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#### **ABSTRACT**

**Objectives.** Sjögren's disease (SjD) is a complex systemic autoimmune disorder with a multistep pathogenesis driven by the interplay of genetic, epigenetic, hormonal, and environmental factors. Even though genetic predisposition is clearly associated with disease susceptibility, it cannot fully explain its pathophysiology. Hormonal fluctuations and various components of the exposome are also considered critical contributors to disease onset and progression. Environmental exposures can induce heritable changes that modulate gene expression without affecting the underlying DNA sequence, known as epigenetic alterations.

Methods. A comprehensive literature search was conducted across major databases (PubMed, MEDLINE, Embase and Web of Science). No publication date restrictions were applied, but only articles written in English were considered. Studies were eligible for inclusion if they investigated any environmental exposure in association with epigenetic modifications in SjD, using human, animal (e.g., mice), or in vitro/ in vivo models.

Results. Epigenetic mechanisms, including DNA methylation, histone modifications, and dysregulation of non-coding RNAs (nc-RNAs), are increasingly recognised as central to SjD pathogenesis. These alterations may serve as a crucial link between the exposome and the onset of autoimmune processes by mediating the immune system's response to external stimuli. Consequently, they may promote aberrant immune activation, leading to both glandular and extra-glandular manifestations, and may increase the risk of lymphomagenesis through chronic Bcell stimulation within affected issues.

Conclusions. Despite notable advances, further research is warranted to elucidate how specific environmental and epigenetic factors interact in genetically susceptible individuals. This systematic review also highlights the potential of epigenetic modifications as biomarkers to enhance diagnostic accuracy and design targeted, individualised therapies based on precision medicine approaches.

#### Introduction

Sjögren's disease (SjD) is a chronic systemic autoimmune disorder typified by lymphocytic infiltration and dysfunction of the salivary and lacrimal glands (1). The disease frequently shows a multifaceted clinical presentation. It may be asymptomatic, especially in early stages, or present with symptoms of xerostomia and xerophthalmia known as sicca symptoms. Additionally, SjD may also manifest with a broad spectrum of extra-glandular and systemic features, potentially involving the joints, skin, kidneys, lungs and other internal organs. Furthermore, SiD is associated with a higher risk of lymphomagenesis in comparison with other autoimmune diseases, probably due to chronic stimulation of B-cells within the affected tissues (2).

Although its exact aetiopathogenesis remains unclear, SjD is widely recognised as a multistep process triggered by environmental, hormonal and epigenetic factors in genetically predisposed individuals, requiring sustained interactions between the innate and adaptive immune systems (3-5). Several genetic susceptibility regions, mainly within but also outside of the major histocompatibility complex (MHC) -including human leukocyte antigen (HLA) and non-HLA genes- have been

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associated with SiD (6). However, genetic predisposition alone, even if different genetic regions are combined in polygenic scores, appears to be insufficient for disease onset and progression (7, 8). Indeed, no absolute concordance rate for SjD in monozygotic twins has been reported, whereas estimates based on different studies for overlapping autoimmune disorders such as systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA) suggest a concordance rate of approximately 20 percent (20%). These findings emphasise the importance of additional cofactors, especially environmental, hormonal and epigenetic influences, in the pathogenesis of SjD.

Among these, epigenetic modifications, particularly those driven by environmental exposures, are thought to play a substantial role in disease pathophysiology (9, 10). By modulating intracellular signalling pathways, they may contribute to aberrant activation of lymphocytes within glandular or extra-glandular tissues, promoting the release of cytokines, such as interferon (IFN)-gamma, interleukin-17 (IL-17), B-cell activating factor (BAFF), as well as the production of SjD specific autoantibodies (anti-Ro/SSA, anti-La/ SSB) (11-13). Each epigenetic mechanism exerts a distinct influence on immune dysregulation in SiD, leading to the activation of diverse molecular pathways and the emergence of phenotypic heterogeneity observed among the patients.

This review provides an overview of recent literature focusing on new insights into epigenetic studies and underscoring how components of the exposome -including infections, diet, smoking, alcohol, pollutants and hormonal fluctuations- may influence gene expression and immune regulation in the context of SjD.

#### Methods

Electronic searches were performed across major databases: PubMed, MEDLINE, Embase and Web of Science. No restrictions were applied regarding publication date, but only articles written in English were considered. The following search string was used

and adapted appropriately for each database: ("Sjogren" OR "Sjogren's" OR "Sjogren's Disease" OR "Sjogren's Syndrome" OR "Sjögren" "Sjögren's" OR "Sjögren's Syndrome" OR "SiD") AND ("environment" OR "environmental factors" OR "environmental exposure" OR "alcohol" OR "smoking" OR "pollution" OR "air pollution" OR "solvents" OR "environmental pollutants" OR "PM2.5" OR "PM10" OR "particulate matter" OR "pesticides" OR "toxins" OR "heavy metals" OR "chemical exposure" OR "diet" OR "nutrition" OR "drug-induced" OR "virus" OR "viral" OR "infection" OR "infections" OR "viral infection" OR "viral infections" OR "EBV" OR "Epstein-Barr" OR "Epstein-Barr virus" OR "CMV" OR "Cytomegalovirus" OR "HCV" OR "Hepatitis C" OR "HTLV-1" OR "HSV" OR "Herpes simplex virus" OR "Hormone Replacement Therapy" OR "hormone therapy" OR "HRT" OR "Estrogen Therapy" OR "ERT" OR "Estrogen Replacement Therapy" OR "stress" OR "psychological stress" OR "chronic stress") AND ("epigenetics" OR "epigenetic" OR "epigenetic modification" OR "epigenetic factors" OR "methylation" OR "hypomethylation" OR "hypermethylation" OR "CpGs" OR "CpG" OR "micro-RNA" OR "micro-RNAs" OR "miRNA" OR "miRNAs" OR "non-coding RNA" OR "noncoding RNA" OR "non-coding RNAs" OR "noncoding RNAs" OR "ncRNA" OR "nc-RNA" OR "ncR-NAs" OR "nc-RNAs" OR "histone" OR "histones" OR "histone modification" OR "histone modifications" OR "histone acetylation" OR "histone hyperacetylation" OR "histone hypoacetylation" OR "histone methylation" OR "histone hypermethylation" OR "histone hypomethylation").

Studies were eligible for inclusion if they investigated any environmental exposure in association with epigenetic modifications in SjD, using human, animal (e.g., mice), or in vitro/ in vivo models. Both descriptive (observational) and interventional designs were considered. Articles were excluded if they did not examine epigenetic effects or were not related to

environmental exposure. All retrieved references were initially screened for duplicates. Then, titles and abstracts were assessed for relevance, followed by full-text evaluation of potentially eligible publications. Two independent reviewers performed the screening process, and any disagreements were resolved by consensus. The workflow of the present review is presented in Figure 1.

#### Epigenetics in Sjögren's disease

Recently, researchers have increasingly highlighted the significant influence of epigenetic mechanisms in the development of autoimmune diseases, including SjD (9, 14, 15).

The term "epigenetics" was first introduced by Conrad Waddington in 1942 within the field of developmental biology, to describe the interactions between genes and their products that result in the expression of a phenotype (16, 17). Since then, the concept has been adopted across various disciplines, leading to some ambiguity regarding its precise definition in the literature. In more recent years, however, "epigenetics" has been more specifically defined as the study of heritable changes that alter gene expression and can potentially influence the phenotype without affecting the underlying DNA sequence. Such changes include DNA methylation, histone modifications, notably via methylation or acetylation, and noncoding RNAs (ncRNAs).

These mechanisms constitute semistable alterations -sufficiently stable to maintain cell identity over time, yet dynamic enough to respond to external stimuli (18, 19). Importantly, epigenetic regulation allows for the coordinated control of multiple genes, enabling cells to activate genes that are necessary for their specific function, while silencing those that are not. This modulation helps maintain stable gene expression patterns within each cell type and supports overall tissue homeostasis. Nevertheless, these patterns are not fixed and can change in response to various environmental factors, including infections, smoking, chemical exposure, and others.

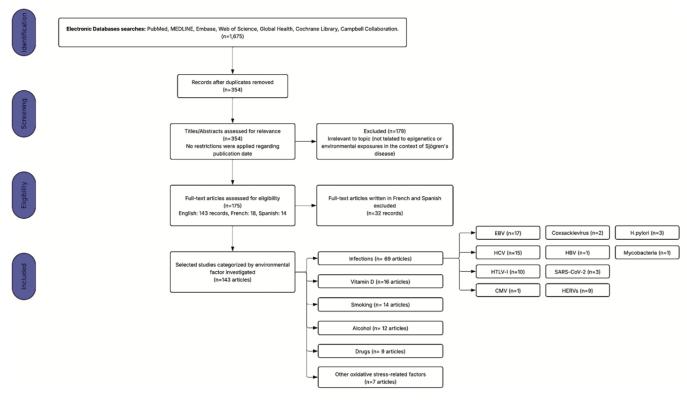


Fig. 1. Workflow of the present review.

#### DNA methylation

The most extensively studied and well-characterised epigenetic mark is DNA methylation. Its primary mechanism involves the inhibition of transcription factors binding to regulatory DNA regions, thereby leading to transcriptional suppression. This process entails the covalent addition of a methyl (CH3) group to cytosine residues within CpG dinucleotides (where a cytosine is followed by a guanine) (19). CpG dinucleotides are not evenly distributed across the genome and tend to cluster in regions known as "CpG islands", which are typically located in gene promoter areas. In somatic cells, promoters of active genes are generally unmethylated, whereas promoters of inactive genes are often hypermethylated (20). In mammalian DNA, methylation takes place almost exclusively at cytosines within CpG contexts and is catalysed by DNA methyltransferases (DNMTs). There are three phylogenetically conserved DNMTs in mammals; DNMT1, DNMT3a and DNMT3b. De novo DNA methylation is established by DNMT3a and DN-MT3b, while DNMT1 maintains these

modifications during DNA replication (20).

Beyond endogenous regulation, DNA methylation patterns can also be modulated by external environmental factors. Specifically, exposure to environmental toxins can directly inhibit DNMTs activity, potentially resulting in hypomethylation or hypermethylation of multiple genes (14). These epigenetic alterations can influence gene expression, with downstream effects on both transcriptional and post-transcriptional levels.

Teruel et al. demonstrated that DNA methylation affects many cell types implicated in SiD pathophysiology, including CD19+ B and CD4+ T lymphocytes, CD14+ monocytes and salivary gland epithelial cells (SGECs) (21). Investigations in these cells from SjD patients employed various methods to assess DNA methylation at specific CpG sites in selected target genes or determined global methylation without single CpG site resolution (19). Epigenome-wide association studies (EWAS) in patients with SjD have consistently revealed a global reduction in DNA methylation levels at CpG sites.

Notably, this hypomethylation was observed within genes involved in type I IFN -such as STAT1, IFI44L, IFITM1 and USP18- signalling across various cell types, particularly in comparison between tissue samples from patients with SjD and healthy individuals. At the molecular level, it was associated with decreased expression of the methyltransferase DNMT1 and increased expression of the demethylating partner Gadd45 alpha (15). Consequently, IFN-inducible genes were upregulated in SiD patients, with a more pronounced signature in individuals exhibiting elevated titres of anti-Ro/SSA and anti-La/SSB autoantibodies (22-24). In addition, CD70 promoter hypomethylation has been shown to lead to overexpression of CD70 in CD4+ T lymphocytes from patients with SiD (25). This epigenetic alteration promotes aberrant T cell activation and enhanced B cell help, ultimately resulting in autoantibody production. Collectively, these findings underscore the role of DNA hypomethylation in disrupting immune tolerance and the broader contribution of epigenetic dysregulation in the pathogenesis and progression of SjD.

#### Histone modifications

Modifications of histone proteins represent another epigenetic mechanism. Histones are small globular proteins (11-15 kD) that participate in the formation of a multi-subunit core around which DNA can be wrapped to create a nucleosome. This structure constitutes the first level of DNA compaction, essential for packaging linear DNA into highly condensed chromosomes, while also being important in gene transcription by modifying chromatin accessibility to the transcriptional machinery (20, 26).

Histones are subject to a wide range of covalent post-translational alterations, among which methylation, acetylation, ubiquitination, phosphorylation being the most prevalent. These modifications have been reported at multiple amino acid residues, although the most common region is the N-terminal tail. As a result, changes of their flexible N-terminal tails can modulate their interactions with adjacent histones, DNA and various nuclear proteins, ultimately impacting gene transcription (27).

The most widely studied histone modifications are the methylation and acetylation of lysine (K) residues, particularly on histones H3 and H4 (28). Histone methylation exerts diverse effects on chromatin structure and cellular processes, including transcriptional activation or repression, as well as involvement in DNA replication and repair. In contrast, histone acetylation is generally associated with a relaxed chromatin conformation that facilitates transcription, whereas deacetylation promotes chromatin condensation and transcriptional repression (28, 29).

Additionally, environmental factors can lead to post-translational histone alterations such as phosphorylation, methylation, or ubiquitination (30). These changes can create binding sites for regulatory proteins that modulate chromatin architecture and gene expression, thereby contributing to the pathogenesis of autoimmune diseases. As reported by Imgenberg-Kreuz *et al.*, CpG sites hypomethylated in whole blood from patients with SjD compared to healthy controls were notably abundant in enhancer regions by histone

3 lysine 4 methylation (H3K4me1) and histone 3 lysine 27 acetylation (H3K27ac), while hypermethylated CpG cites were depleted in these areas and primarily localized within genes marked by histone 3 lysine 36 methylation (H3K36me3), indicative of active transcription (19, 31). At present, there are no studies available analysing histone modifications directly in primary cells from patients with SjD. Nevertheless, efforts to integrate multiple layers of genome-wide datasets, including SNP genotyping, DNA methylation and histone marks, offer significant potential to enhance our understanding of the functional impact of variants associated with SjD.

#### Non-coding RNAs (ncRNAs)

Among the mechanisms of epigenetic regulation are also non-coding RNAs (ncRNAs), a diverse group of RNA molecules that do not encode functional proteins; however, they act as regulatory RNAs, essential for the post-transcriptional control of mRNA expression (32, 33). NcRNAs include microRNAs (miRNAs) and long noncoding RNAs (lncRNAs). Their transcription from intronic or intergenic regions plays a critical role in biological processes such as tissue differentiation, development, proliferation and cell metabolism.

MiRNAs are a class of evolutionary highly conserved small (19-25 nucleotides), non-coding, single-stranded RNA molecules that play a pivotal role in gene silencing by acting as negative modulators of gene expression (34-36). They perform this function by binding to complementary sequences of their target mRNA in the 3' untranslated region (UTR) (36). By this mechanism, miRNAs induce either translational inhibition or destabilization of the target mRNA (37). LncRNAs, on the other hand, are defined as ncRNAs with a length above 200 nucleotides, localized in both the nucleus and the cytoplasm (38, 39). They participate in a range of molecular pathways, including epigenetic regulation, transcription and translation, although their precise mechanisms of action are still elusive. It has been observed that lncRNAs can interact with other ncRNAs, such as miRNAs. Specifically, they may function as "miRNA sponges", binding to miRNAs and thus preventing them from targeting mRNAs (40). Moreover, lncRNAs can compete with the 3' UTR regions of mRNAs for miRNA binding, which can indirectly inhibit the miRNA-mediated repression of these mRNAs (41).

Emerging evidence suggests that dysregulation of ncRNAs may contribute to SjD pathogenesis by modulating immune responses and inflammatory signaling pathways (42). Aberrant expression of both miRNAs and lncR-NAs has been identified across multiple autoimmune disorders, including SiD (37, 43, 44). Indeed, recent studies in peripheral blood cells, minor salivary gland tissue and SGECs from patients with SjD revealed numerous differentially expressed miRNAs (45). Nevertheless, the exact functional implications and target genes of these molecules remain only partially understood in many cases (45-50).

Owing to their participation in fundamental cellular regulation, alterations in expression of ncRNAs can significantly affect their target molecules. One of the most extensively studied and well-characterised miRNAs, hasmiR-146a, is known to have a key role in both innate and adaptive immunity by suppressing inflammatory responses (51). Investigations into the expression profile of this miRNA in peripheral blood mononuclear cells (PBMCs) from individuals with SjD have consistently demonstrated its upregulation (47). Consequently, has-miR-146a is thought to participate in early disease pathogenesis by promoting phagocytic activity and reducing the production of pro-inflammatory cytokines (47).

Additionally, recent findings have shown that downregulation of miR-574 and miR-17-92, along with upregulation of miR-768-3p,miR-150 and miR-155, are also associated with the pathogenesis of SjD (52, 53). Bruno *et al.* observed miR-155 upregulation in SjD patients who experienced non-Hodgkin's lymphoma (NHL), that was correlated positively with focus score, BAFF-R, and IL-6R levels- biomarkers

of aberrant B cell activation and lymphomagenesis (54).

Environmental exposures have been shown to modulate the transcription of miRNAs and lncRNAs through various mechanisms, including direct effects on transcription factor effectiveness, epigenetic modifications or disruptions in intracellular signalling pathways (55). Moreover, the gut microbiota, which is dynamically shaped by diet and other environmental elements, can modulate ncRNA expression, indirectly affecting immune system regulation and the initiation or progression of autoimmune disorders (56, 57).

# **Environmental factors in the pathogenesis of Sjögren's disease**

Viral infections

Viral infections have long been considered potential triggers for initiating the autoimmune process in SjD (58). As a result, they have been the subject of ongoing investigation. Thus far, three different viruses have been extensively studied in relation to their impact on the pathogenic pathways of SiD; Epstein-Barr virus (EBV), hepatitis C virus (HCV) and human T cell lymphotropic virus type I (HTLV-1) (58, 59). However, there are many other viruses that are thought to be associated with SjD development, which will be further examined below (60-63). The mechanisms by which viruses can induce modifications in infected cells are still debated but may involve abnormal gene transcription resulting from viral genome insertion into the host cell DNA, or virus-induced epigenetic alterations.

Epstein-Barr virus (EBV). Although it should be noted that EBV is highly prevalent, being present in up to 95% of the general population, it appears to exhibit a marked tropism for the salivary glands, suggesting a potential association with SjD. Indeed, several studies have shown that EBV DNA is detected at increased levels in the salivary glands of patients with SjD compared with healthy individuals. Moreover, patients positive for anti-SSA/Ro and/or anti-SSB/La autoanti-bodies have been found to have higher

immunoglobulin G (IgG) titres against the EBV early antigen in contrast to those with negative autoantibodies. On the same prism, Hudson *et al.* studied previously healthy females with primary EBV infection and concluded that anti-Ro52 and anti-Ro60 autoantibodies were detectable as early as seven days after infection, undergoing class switching from IgM to IgG (64). These findings suggest that EBV infection may directly promote the production of SjD-associated autoantibodies.

Recently, Kaberdoos et al. found that genes such as SLAM7, RUNX3, BCL2, FAS and CD247, which are involved in the immune response to EBV, were observed to be hypomethylated in labial salivary gland (LSG) tissue of patients with SiD (65). Specifically, SLAM7 has been identified as a marker expressed on CD4+ cytotoxic T lymphocytes (CTLs). In SjD, elevated levels of CD4+ CTLs have been reported in both peripheral blood and SGECs, correlating with disease progression and severity (66). Notably, CD4+ CTLs comprise approximately 20% of all T lymphocytes in the salivary glands of affected individuals. Recent investigations proved that increased expression and hypomethylation of the RUNX3 gene in minor salivary gland tissue have demonstrated excellent diagnostic performance with an area under the curve (AUC) of 1.0 for distinguishing SiD patients (67). Additionally, overexpression of BCL2, FAS and FASL in LSG of SjD has been confirmed by immunohistochemistry, with BCL2 upregulation correlating with a reduction in acinar cell numbers (68). Complementary findings from genome-wide DNA methylation analyses reported CD247 hypomethylation in CD4+ T lymphocytes of SjD patients, suggesting epigenetic regulation may contribute to disease pathogenesis (8, 22, 69). The observed increase in B-cell populations and hypomethylation of EBVrelated genes underscore their involvement in the development of ectopic lymphoid structures within the salivary glands of SjD patients and, consequently, in disease pathogenesis (70). Importantly, EBV is detectable within these structures in a subset of patients,

whereas it is absent in glandular tissue lacking these structures.

As previously mentioned, miRNAs expression plays also a central role in SiD pathogenesis. One study reported that levels of the EBV-derived miRNA, ebv-miR-BART13-3p, were elevated in the salivary glands of patients with SiD compared to healthy controls (71). Of note, ebv-miR-BART13-3p has been proposed to suppress the expression of stromal interaction molecule 1 (STIM1), a key regulator in the secretory function of acinar cells. This finding provides a compelling mechanistic link between EBV infection and reduced saliva production. Furthermore, this miRNA has been reported to mediate intercellular communication by transferring between B lymphocytes -the primary targets of EBV- and SGECs via microvesicles.

Moreover, latent transcripts of EBV [such as EBV nuclear antigens (EB-NAs) and latent membrane proteins (LMPs)] exert epigenetic control over viral oncoprotein expression through mechanisms including CpG methylation, histone deacetylation, miRNA interactions, and activation of super enhancers (72). These same epigenetic pathways may also lead to the immortalisation of host cells and to abnormal cell behaviour that might result in malignancy. In addition, EBV encodes two ncRNAs, EBER-1 and EBER-2, which have been associated with B cell lymphomagenesis. These findings suggest that EBV may influence the SjD pathogenesis through epigenetic reprogramming of B lymphocytes, resulting in both chronic immune activation and lymphoproliferative disorders.

Hepatitis C virus (HCV). HCV has been increasingly implicated in autoimmune manifestations that resemble or potentially overlap with SjD (73, 74). In addition to its well-established hepatotropism, it exhibits both lymphotropism and sialotropism, infecting immune cells and salivary glands -tissues central to SjD pathology (75).

Chronic HCV infection has been associated with systemic immune activation, particularly B-cell hyperactivity mediated by elevated BAFF levels,

which may lead to features commonly observed in SjD, such as cryoglobulinemia, vasculitis, polyneuropathy and lymphoproliferative disorders (76, 77). These extrahepatic manifestations, alongside clinical symptoms like xerostomia and arthralgia, have led to ongoing debate over whether HCV acts as a direct etiological factor in SjD or simply mimics its clinical presentation. Nonetheless, it should be noted that active HCV infection is an exclusion criterion in the 2016 American College of Rheumatology/European League Against Rheumatism (currently European Alliance of Associations for Rheumatology) classification criteria for SjD (78).

Recent findings suggest that HCV may exert epigenetic effects on host cells, possibly altering gene expression profiles involved in immune pathways and epithelial cells function, further reinforcing its potential relevance in SjD pathogenesis (79). Although such epigenetic marks, namely DNA methylation, histone modifications and miRNA dysregulation, have been documented in HCV-infected hepatocytes and lymphoid cells, direct evidence of these alterations in SGECs remains lacking (80).

Human T Lymphotropic virus type I (HTLV-I). Investigations using transgenic mouse models have demonstrated that retroviral infection increases susceptibility to autoimmune diseases, including SjD, polymyositis and RA, compared to non-infected controls (81). In this context, HTLV-I, through the expression of its regulatory Tax and HBZ (HTLV-I bZIP factor), can infect SGECs, resulting in elevated levels of proinflammatory mediators, such as ICAM-1, IP-10, and chemokines like RANTES (Regulated on Activation, Normal T cell Expressed and Secreted) (82). During HTLV-I infection, B cell activity appears to be downregulated, leading to reduced production of autoreactive antibodies (83).

Green *et al.* reported that transgenic mice with the HTLV-1 Tax gene developed an exocrinopathy resembling human SjD (84). Additionally, Mariette *et al.* identified the HTLV-I Tax gene

within salivary gland cells of SjD patients (85). Nakamura et al. observed that SjD patients co-infected with HTLV-I and HAM had significantly lower anti-SAA/Ro antibody titres, despite more pronounced lacrimal gland involvement; however, Focus scores in minor salivary gland biopsies were comparable between HTLV-I-positive and -negative groups (83). Terada et al. demonstrated that HTLV-I-seropositive SjD patients often have IgAclass HTLV-I antibodies in saliva (86). In follow-up research, the same group noted reduced glandular destruction in HTLV-I-positive SjD patients, suggesting that HTLV-I may inhibit apoptotic pathways and promote cell proliferation (87).

A Brazilian cohort of 129 HTLV-I-seropositive individuals showed that, although many exhibited sicca (46 with xerostomia, 18 with xerophthalmia, 8 with confirmed hyposalivation), only one had SjD-specific autoantibodies; minor salivary gland biopsies in six cases showed SjD-characteristic mononuclear infiltrates (88).

Overall, these findings suggest a dual role for HTLV-I; in some individuals, it may trigger a full-blown autoimmune response culminating in SjD, while in others it may cause nonspecific salivary gland inflammation manifesting as sicca symptoms. Notably, HTLV-Ipositive SjD patients tend to have fewer ectopic germinal centres and reduced CXCL13 expression in infiltrating mononuclear cells in contrast to HTLV-I-negative SjD patients (89). Nevertheless, according to the current literature, there is no evidence to date indicating that HTLV-I induces epigenetic modifications in SGECs. Therefore, more indepth investigation is warranted in this context of SjD pathogenesis.

#### Other viruses

As previously stated, several other viruses have been implicated in the potential pathogenesis of SjD, including, among others, cytomegalovirus (CMV), coxsackievirus, hepatitis B virus (HBV) and SARS-CoV-2 virus. The salivary glands are recognised as target organs of CMV, with cases of parotitis and sialadenitis being described during

the course of infection (90). Triantafyllopoulou et al. detected evidence of coxsackie virus VP1 antigen in minor SGECs of SjD patients, an infection not observed in secondary SjD, other rheumatic conditions or healthy controls (60). The authors also reported cross-reactivity between anti-SAA/ Ro60 and a homologous peptide in coxsackievirus 2B protein, suggesting virus-induced immune activation (61). Additionally, HBV has been associated with SjD through observational data indicating a potential protective effect of antiviral therapy (62). Despite these findings, current evidence is insufficient to establish a clear link between these viral infections and SjD development. This field remains unexplored, and it is currently unknown whether and how these viruses may contribute to SiD pathophysiology, particularly via epigenetic mechanisms.

According to current studies, the prevalence of Coronavirus disease-19 (COVID-19) has been found to be significantly higher in patients with SiD compared to general population. In the study by Lee et al., the comparison between COVID-19-positive and COVID-19-negative individuals identified three differentially methylated CpG sites with a false discovery rate (FDR) below 0.05: cg22399236, cg03607951, and cg09829636 (91). Notably, a search of these CpGs in the EWAS Atlas revealed multiple associations between hypomethylation of cg03607951, located in IFI44L gene, and several autoimmune disorders, including SLE, SjD, and mixed connective tissue disease, among others. The persistence hypomethylation of IFI44L three months post-infection may indicate that the reversal of infectioninduced immune responses is a prolonged process, potentially reflecting the lasting impact of SARS-CoV-2 on the epigenetic regulation of immunerelated genes. In this context, longitudinal studies assessing the persistence of methylation changes after infection could provide valuable insights into the possible long-term epigenetic consequences of COVID-19 and their role in the development of autoimmune diseases (63, 69).

Human endogenous retroviruses (HERVs). HERVs, which comprise approximately 8% of the human genome, are normally silenced by epigenetic mechanisms, primarily DNA methylation (92). Disruption of this regulation can lead to the expression of HERV elements and retroviral proteins with immunogenic potential. In SLE and SjD, specific HERV loci, such as HRES-1, HERV3-1 and HERV-E 4.1 p30 gag protein, are overexpressed in lymphocytes and SGECs, and antibodies against HRES-1-derived p38 gag protein have been found in a subset of patients with both autoimmune diseases (93). Furthermore, DNA hypomethylation of HERV-CD5 promoter in B lymphocytes results in the expression of an intracellular CD5 variant, which may drive autoreactive responses (94, 95). At the molecular level, both SLE and SiD share a defective protein kinase (PKC)-delta/ERK/DNA methyltransferase (DNMT)1 pathway -affecting lymphocytes in SLE and SGECs in SiD- which leads to global DNA hypomethylation (96). Histone modifications are also detected in CD4 T cells from SLE patients, with global H3 and H4 hypoacetylation and hyper H3k9 trimethylation (97). Methylation inhibitors such as 5-azacytidine (5-aza-C) or procainamide seem to enhance HERV transcription by downregulating DNMT1, a phenomenon observed both in PBMCs and in animal models. Moreover, experimental agents, including cycloheximide (a protein synthesis inhibitor) and benzopyrene diol epoxide (a DNA-damaging compound), have been shown to synergistically upregulate ERV RNA expression in vitro. Remarkably, the epigenetic dysregulation underlying HERV activation appears to be reversible. In SjD, treatment with anti-CD20 monoclonal antibody (rituximab) has been shown to restore methylation profiles in SGECs and correct pathway defects (96). Similarly, in SLE, anti-IL-6 receptor therapy (tocilizumab) reactivates DNMT1 expression through the ERK pathway in B lymphocytes (98, 99). These findings strongly support a model in which epigenetically driven HERV dysregulation contributes to the pathogenesis

of autoimmune diseases like SLE and SjD, particularly in genetically predisposed individuals.

#### Other infections

Helicobacter pylori. H.pylori infection may play a role in SiD pathogenesis; however, its contribution remains controversial due to conflicting reports (100). A recent meta-analysis of nine studies examining H.pylori infection rates among SiD patients reported a weak association between the two entities, reaching an odds ratio (OR) of 1.24 (95% CI 1.03-1.50) (101). Notably, six out of the nine included studies used outdated classification criteria for SjD, while only two confirmed H.pylori infection via biopsy; the remaining seven relied on serological testing. Therefore, the findings from this meta-analysis should be interpreted with caution. Additionally, a more recent investigation showed a higher prevalence of past H.pylori infection among newly diagnosed SiD patients, compared to controls.

MiRNA-mediated immune modulation and impaired apoptosis observed in H.pylori-associated gastric MALT lymphoma raise the possibility of similar epigenetic dysregulation in SjD. Specifically, H.pylori infection appears to affect the expression of certain miRNAs, such as 21a, 135b, 142a, 150, and 155, which modulate cell proliferation, B cell hyperactivity, and survival. These miRNAs have been shown to inhibit the transcription of the gene encoding the proapoptotic protein TP53INP1 (tumour protein p53-inducible nuclear protein 1) (102). Thus, it is plausible that H.pylori, through epigenetic alterations, may be involved in SjD pathogenesis and potentially lead to complications such as lymphoma.

Mycobacterial infection. Recent findings suggest that infection with non-tuberculosis mycobacteria (NTM), such as Mycobacterium avium subsp. paratuberculosis (MAP), may contribute to the pathogenesis of SjD through mechanisms involving molecular mimicry and epigenetic modulation of immune responses. In genetically

predisposed individuals, including those carrying polymorphisms in the TNFAIP3 gene, which encodes the A20 protein, dysfunction of A20 may amplify macrophage inflammatory responses to mycobacteria, increasing exposure to mycobacterial antigens. MAP, widely present in the environment and the diet, expresses heat shock proteins (e.g., hsp65) that share epitope homology with host autoantigens Ro and La, supporting a molecular mimicry mechanism in autoimmunity. Moreover, the BCG vaccine -known to modulate both innate and adaptive immunity via epigenetic reprogramming- has been proposed to exert protective or therapeutic effects against SjD, underscoring a potential role of infection-driven epigenetic alterations in the initiation or development of the disease (103).

#### Vitamin D

In SjD, several studies have suggested a potential role for vitamin D in the disease pathogenesis (104-106). The primary natural source of vitamin D (1,25-dihydroxyvitamin D3) is its synthesis in the skin, which occurs through a process dependent on ultraviolet radiation (UV). Reduced exposure to UV, often recommended as part of the management of cutaneous manifestations, has been proposed as a contributing factor to vitamin D deficiency (107). Recently, increasing attention has been given to the role of vitamin D in modulating epigenetic mechanisms, primarily in the context of cancer, as well as in the pathogenesis of autoimmune disorders (108, 109). Among these, SLE is the most extensively studied in relation to epigenetic dysregulation, with vitamin D deficiency being implicated in disrupted DNA methylation -particularly through reduced availability of methyl donors and impaired DNMT1 activity- leading to gene hypomethylation and immune imbalance (112). Furthermore, focus has been placed on the methylation status of specific genes in T lymphocytes, such as CD11a (ITGAL), perforin (PRF1), CD70 (TNFSF7), and CD40LG (TNFSF5), as key contributors to the pathogenesis and progression of SLE (14). Histone modifications and ncRNAs in SLE have received less research attention compared to DNA methylation.

In contrast, in RA, vitamin D deficiency has been linked to higher disease activity, and certain polymorphisms and methylation changes in vitamin D genes (such as VDR and CYP27B1) have been associated with disease susceptibility.

In SjD, although low vitamin D levels have been correlated with disease manifestations and complications, evidence for a direct epigenetic mechanism remains limited. Conflicting results from other studies underscore the need for further investigation. Of note, a vitamin D-related SNP, VDR rs7975232, has been proposed as a potential risk factor for SiD, although its relevance appears to be population-specific (113). Therefore, additional largescale, multi-ethnic genetic studies are urgently warranted to unravel the interaction between vitamin D, genetic susceptibility and disease progression. Comparable limitations are observed in systemic sclerosis and inflammatory bowel disease, where vitamin D status influences immune function and epithelial barrier integrity, but its role in epigenetic regulation is not yet clearly defined (114, 115).

#### Smoking

Despite its well-established role as a risk factor in autoimmune diseases, such as RA and multiple sclerosis, the impact of smoking on SjD remains insufficiently investigated. Nevertheless, Jin et al. (116), in their systematic review and meta-analysis on risk factors for SjD, examined the association between smoking and SjD in detail. According to their findings, current smoking was negatively associated with the development of SjD, with all reported ORs being less than 1 (<1) (117-120). Although several studies have suggested that smoking may have a protective effect against SjD (59, 119), Jin et al. did not reveal any clear association between either former or current smoking and the disease (116). Of note, Olsson et al. reported that individuals who later developed SjD smoked at similar levels during early life as the general

population but were more likely to quit smoking over time (117).

Cigarette smoking leads to the inhalation of toxic substances, including tars, nicotine, carbon monoxide, polycyclic aromatic hydrocarbons, and free radicals. This toxic exposure induces oxidative stress, which can disrupt normal epigenetic regulation. One proposed mechanism involves the inhibition of MEK-ERK kinase signalling pathway, which may alter gene expression in immune and inflammatory pathways, thereby contributing to the pathogenesis of autoimmune diseases (123-125). Inhibition at any point in this molecular pathway may ultimately result in the downregulation of DNMTs. The consequent DNA hypomethylation can lead to the upregulation of pro-inflammatory genes, potentially driving clinical manifestations that resemble those observed in SLE (126).

These alterations may also affect additional components of the ERK signalling pathway, as well as its regulatory proteins, including specific phosphatases. For example, protein phosphatase 2A (PP2A) directly inactivates MEK, leading to inhibition of ERK. Other phosphatases involved include PPS and DUSP23, the latter of which being notably overexpressed in patients with SLE. Furthermore, several proteins can modulate the activity of DNMTs. One such protein is EZH2, the catalytic subunit of the Polycomb Repressive Complex 2; elevated EZH2 expression in CD4+ T lymphocytes has been associated with DNMT3A repression, resulting in epigenetic dysregulation in autoimmune disorders (127).

There is evidence suggesting that some of these methylation modifications may be reversible following to-bacco cessation (125, 128). However, the interplay between gene polymorphisms, epigenetic marks, and smoking exposure remains incompletely understood and requires further investigation.

As mentioned above, such epigenetic influences have been observed in SLE, where smoking-related alterations in DNA methylation have been linked to disease susceptibility and severity

(129, 130). This raises the possibility that similar mechanisms may also be involved in SjD pathogenesis, indicating a need for further research to explore whether tobacco-induced epigenetic changes contribute to disease onset or progression in SjD as well.

#### Alcohol

Alcohol consumption has been shown to induce epigenetic modifications, including DNA methylation and histone acetylation (131). These alterations can disrupt immune homeostasis and may contribute to the initiation or progression of autoimmune disorders (132). Although most existing data focus on SLE, similar epigenetic pathways are thought to be involved in SjD, which shares overlapping immunopathogenic mechanisms (133, 134). Antioxidant compounds found in alcoholic beverages, most notably resveratrol in wine and humulones in beer, may further modulate these epigenetic processes, possibly inhibiting inflammatory pathways (135-137). Moreover, interindividual variability in alcohol metabolism, such as polymorphisms in genes like N-acetyltransferase 2 (NAT2), has been associated with altered risk for SLE (138).

Given the shared molecular features between SLE and SjD, including T cell DNA hypomethylation and IFN-regulated gene signatures, it is plausible that alcohol-induced epigenetic alterations -modulated by metabolic genotypes like NAT2- could similarly affect disease susceptibility or activity in SjD. Future research integrating epigenomic and pharmacogenomic data is needed to elucidate these interactions and their potential relevance in disease modulation or prevention.

#### Drug-induced SjD

Hydralazine, procainamide and isoniazid have been implicated in the development of autoimmune responses through their capacity to disrupt normal epigenetic regulation in T lymphocytes (139-141). Both agents inhibit DNA methylation, particularly in CD4<sup>+</sup> T lymphocytes, promoting the expression of normally silenced immune-related genes. *In vitro* studies

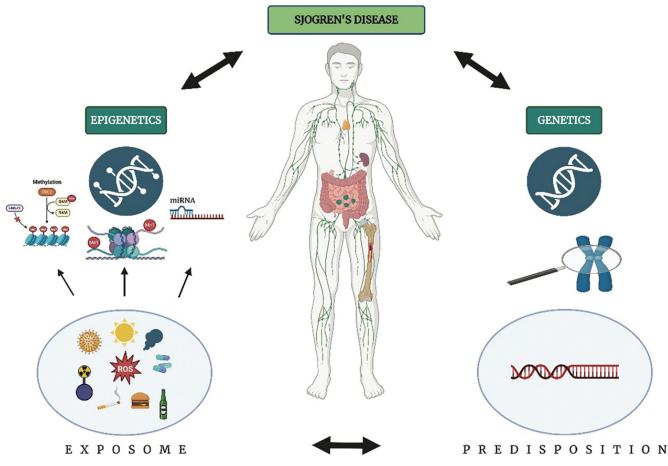


Fig. 2. Genetics and epigenetics in Sjögren's disease (SjD).

demonstrate that exposure of human T cell clones to these drugs results in autoreactive phenotypes (142). Similarly, murine models receiving CD4+ T lymphocytes treated with these drugs develop autoimmune manifestations, including glomerulonephritis and production of autoantibodies such as anti-dsDNA and anti-histone antibodies (130).

Mechanistic investigations reveal that these drugs reduce DNMT1 activity and lead to overexpression of genes such LFA-1, CD70, and CD40L. These changes have been observed both in patients with SLE and in drug-induced lupus animal models, underscoring a shared pathogenic mechanism (143). Considering the similarities in DNA methylation alterations reported in both SLE and SjD, notably the hypomethylation of IFN-stimulated genes and CD70, emerging recent studies suggest that comparable epigenetic modifications may play a role in the onset or progression of SiD.

Oxidative stress

Oxidative stress represents a key environmental factor that influences gene expression through epigenetic mechanisms and plays a pivotal role in the pathogenesis of autoimmune diseases, including SLE and SjD.

In addition to smoking, various environmental exposures contribute to oxidative stress (144). These include, among others, diet, UV radiation (especially UVB; 290-320 nm) (113), air pollution (e.g., nitrogen dioxide, ozone), industrial solvents, pesticides, silica dust, and toxic heavy metals like cadmium and arsenic. All these factors promote the excessive production of reactive oxygen species (ROS), leading to oxidative damage of cellular components and disruption of intracellular signalling pathways.

Importantly, oxidative stress also affects the function of epigenetic enzymes, especially DNMT1. Reduced DNMT1 activity results in global DNA hypomethylation, particularly in Tlym-

phocytes, allowing aberrant expression of normally silenced immune-related genes. This process may enhance immune hyperactivity and cause the loss of self-tolerance, leading to lupus-like disease in genetically predisposed individuals (140, 145, 146).

Moreover, certain dietary components, such as folate and choline, constitute essential methyl donors for DNA methylation. Insufficient intake or impaired metabolism of these nutrients can disrupt the biochemical supply of methyl groups necessary for DNMT function, thereby promoting aberrant DNA methylation patterns (147). A diet poor in methyl donors may aggravate DNA hypomethylation caused by oxidative stress, leading to immune dysregulation and potentially contributing to the development of autoimmune disorders, including SLE and SjD.

A recent genome-wide methylation analysis of SLE patients identified an association between living near major highways, an indicator of long-term

## Epigenetic effects of the exposome in Sjögren's disease / E.D. Prifti et al. $\,$

Table I. Environmental factors and their epigenetic influences in Sjögren's disease (SjD) and other autoimmune conditions.

	1 6 - J 6	( ) /	
	DNA methylatio	on	
Environmental factor	Main findings in SjD	Main findings in other autoimmune conditions	References
Epstein-Barr virus (EBV)	-RUNX3 hypomethylation in minor salivary glands -CD247 hypomethylation in CD4+ T lymphocytes		(8, 22, 65, 67)
SARS-CoV-2 virus	Hypomethylation of cg03607951 in IFI44L gene	SLE, mixed connective tissue disease Hypomethylation of cg03607951 in IFI44L gene	(63, 69, 92)
HERVs	-DNA hypomethylation of HERV-CD5 promoter in B lymphocytes -Defective PKC-δ/ERK/DNMT1 & global DNA hypomethylation -Reversibility (rituximab)	SLE -Defective PKC-δ/ERK/DNMT1 & global DNA hypomethylation -Reversibility (tocilizumab)	(95, 96, 97, 99, 100)
1,25-dihydroxyvitamin D3 & Methyl-donor-poor diet	-Impaired DNMT1 activity - VDR rs7975232 as a potential risk factor	SLE -Impaired DNMT1 activity -Hypomethylation of CD11a (ITGAL), PRF1, CD70 (TNFSF7) and CD40LG (TNFSF5) in T lymphocytes RA Methylation changes in vitamin D genes (including VDR & CYP27B1) and disease susceptibility	(14, 112-115, 147, 151)
Smoking	-Oxidative stress -Inhibition of MEK-ERK kinase signalling pathway -Downregulation of DNMTs & DNA hypomethylation -Upregulation of pro-inflammatory genes -Potential association with SjD susceptibility and severity -Need for further research in SjD pathogenesis	SLE -Oxidative stress -Inhibition of MEK-ERK kinase signalling pathway -Downregulation of DNMTs & DNA hypomethylation -Upregulation of pro-inflammatory genes -PPS & DUSP23 overexpression -Association of EZH2 overexpression in CD4+ T lymphocytes with DNMT3 inhibition	(123-125) (126) (127) (128) (129, 130)
Hydralazine, procainamide, isoniazid (INH)	-Downregulation of DNMT1 -DNA hypomethylation of IFN-stimulated genes in CD4+ T lymphocytes -Overexpression of LFA-1, CD70, and CD40L genes	SLE -Downregulation of DNMT1 -DNA hypomethylation of IFN-stimulated genes in CD4+ T lymphocytes -Overexpression of LFA-1, CD70, and CD40L genes	(94, 130, 139-141, 142, 152
Oxidative stress: -UV radiation -Air pollution -Industrial solvents -Pesticides -Silica dust -Heavy metals (e.g., cadmium, arsenic)	-Excessive production of ROS -Reduced DNMT1 activity -Global DNA hypomethylation, particularly in T lymphocytes	SLE -Excessive production of ROS -Reduced DNMT1 activity -Global DNA hypomethylation, particularly in T lymphocytes -Association of long-term exposure to traffic-related air pollution with UBE2U gene hypomethylation	(140, 145, 146, 148)
	Histone modificati	ions	
Environmental factor	Main findings in SjD	Main findings in other conditions	References
Epstein-Barr virus (EBV)		EBNAs & LMPs, histone deacetylation and activation of super enhancers (B cell lymphomagenesis)	(72)
HERVs	-Global H3 & H4 hypoacetylation -Hyper H3k9 trimethylation	-	(98)
	Non-coding RNAs (no	-RNAs)	
Environmental factor	Main findings in SjD	Main findings in other conditions	References
Epstein-Barr virus (EBV)	Elevation of EBV-miR-BART13-3p in salivary glands of SjD patients	Association of EBER-1 and EBER-2 ncRNAs with B cell lymphomagenesis	(71)
Helicobacter pylori	Dysregulation of miR-21a, miR-135b, miR-142a, miR-150, miR-155 Inhibition of TP53INP1		(103)

exposure to traffic-related air pollution, and hypomethylation of the UBE2U gene (148). This gene encodes an enzyme involved in protein and histone ubiquitination, as well as DNA repair, suggesting the environmental pollutants may influence epigenetic regulation of key cellular pathways. Further studies are needed to confirm these findings related specifically to SLE patients. Similar research is also warranted in SjD in order to determine whether environmental exposures such as air pollution induce comparable epigenetic alterations.

#### **Conclusive remarks**

Epigenetics is emerging as a crucial area of research in SjD, offering significant insights into the underlying mechanisms of the disease. By revealing how dynamic and potentially reversible modifications regulate gene expression, epigenetic studies open promising avenues for clinical innovation. These alterations hold potential not only as diagnostic biomarkers but also therapeutic targets, creating new opportunities for precision medicine approaches in the treatment of SjD (149, 150).

Recent studies have identified epigenetic changes in both immune cells and SGECs affected by SiD, emphasising their key role in disease pathophysiology through the dysregulation of intracellular signalling pathways. As a consequence, they may drive immune dysfunction and result in glandular damage. Moreover, various components of the exposome, such as viral infections, smoking, alcohol consumption, and exposure to pollutants or toxins, appear to influence these epigenetic mechanisms, potentially contributing to the onset and progression of autoimmune diseases, including SiD.

Despite notable progress, substantial investigation is still needed to clarify how specific epigenetic and environmental factors interact in genetically predisposed individuals, leading to SjD development and heterogeneity. Continued exploration in this field could significantly improve diagnostic accuracy, prognostic assessment, and therapeutic precision in SjD

through individualised, mechanism-driven strategies.

A summary of the epigenetic modifications influenced by specific environmental factors in SjD and other autoimmune conditions is presented in Table I.

#### References

- MARIETTE X, CRISWELL LA: Primary Sjögren's Syndrome. N Engl J Med 2018; 378(10): 931-9.
  - https://doi.org/10.1056/NEJMcp1702514
- CHATZIS LG, STERGIOU IE, GOULES AV et al.: Clinical picture, outcome and predictive factors of lymphoma in primary Sj gren's syndrome: results from a harmonized dataset (1981-2021). Rheumatology (Oxford) 2022; 61(9): 3576-85. https://doi.org/10.1093/rheumatology/keab939
- 3. FOX RI: Sjögren's syndrome. *Lancet* 2005; 366(9482): 321-31. https://doi.org/10.1016/S0140-6736(05)66990-5
- CHATZIS L, VLACHOYIANNOPOULOS PG, TZIOUFAS AG, GOULES AV: New frontiers in precision medicine for Sjögren's syndrome. Expert Rev Clin Immunol 2021; 17(2): 127-41. https://
  - doi.org/10.1080/1744666X.2021.1879641
- KARAGIANNI P, TZIOUFAS AG: Epigenetic perspectives on systemic autoimmune disease. J Autoimmun 2019; 104: 102315. https://doi.org/10.1016/j.jaut.2019.102315
- TEOS LY, ALEVIZOS I: Genetics of Sjögren's syndrome. Clin Immunol 2017; 182: 41-7. https://doi.org/10.1016/j.clim.2017.04.018
- FUGMANN C, REID S, PUCHOLT P et al.: A high polygenic risk score is associated with SSA/SSB antibody positivity and early onset in primary Sjögren's disease. Rheumatology (Oxford) 2025; 64(7): 4341-6. https:// doi.org/10.1093/rheumatology/keae693
- KHATRI B, TESSNEER KL, RASMUSSEN A et al.: Genome-wide association study identifies Sjögren's risk loci with functional implications in immune and glandular cells. Nat Commun 2022; 13(1): 4287. https://doi.org/10.1038/s41467-022-30773-y
- 9. LU Q: The critical importance of epigenetics in autoimmunity. *J Autoimmun* 2013; 41: 1-5. https://doi.org/10.1016/j.jaut.2013.01.010
- JONSSON R, BROKSTAD KA, JONSSON MV, DELALEU N, SKARSTEIN K: Current concepts on Sjögren's syndrome - classification criteria and biomarkers. *Eur J Oral Sci* 2018; 126 Suppl 1(Suppl Suppl 1): 37-48. https://doi.org/10.1111/eos.12536
- LIP, HAN M, ZHAO X, REN G, MEI S, ZHONG
   C: Abnormal Epigenetic Regulations in
   the Immunocytes of Sjögren's syndrome
   Patients and Therapeutic Potentials. Cells
   2022; 11(11).
   https://doi.org/10.3390/cells11111767
- PERRICONE C, BRUNO L, CAFARO G et al.: Sjögren's syndrome: Everything you always wanted to know about genetic and epigenetic factors. Autoimmun Rev 2024; 23(12): 103673.
- https://doi.org/10.1016/j.autrev.2024.103673 13. CHATZIS LG, GOULES AV, TZIOUFAS AG:

- Searching for the "X factor" in Sjögren's syndrome female predilection. *Clin Exp Rheumatol* 2021; 39 (Suppl. 133): S206-14. https://
- doi.org/10.55563/clinexprheumatol/88dyrn
   DANIELI MG, CASCIARO M, PALADINI A et al.: Exposome: Epigenetics and autoimmune diseases. Autoimmun Rev 2024; 23(6): 103584.
  - https://doi.org/10.1016/j.autrev.2024.103584
- JEFFRIES MA, SAWALHA AH: Autoimmune disease in the epigenetic era: how has epigenetics changed our understanding of disease and how can we expect the field to evolve? Expert Rev Clin Immunol 2015; 11(1): 45-58. https://
- doi.org/10.1586/1744666X.2015.994507
- HOLLIDAY R: Epigenetics: an overview. *Dev Genet* 1994; 15(6): 453-7. https://doi.org/10.1002/dvg.1020150602
- NOBLE D: Conrad Waddington and the origin of epigenetics. *J Exp Biol* 2015; 218(Pt 6): 816-8.
   https://doi.org/10.1242/jeb.120071
- SCHÜBELER D: Function and information content of DNA methylation. *Nature* 2015; 517(7534): 321-6.
  - https://doi.org/10.1038/nature14192
- IMGENBERG-KREUZ J, SANDLING JK, NOR-DMARK G: Epigenetic alterations in primary Sjögren's syndrome - an overview. *Clin Immunol* 2018; 196: 12-20.
- https://doi.org/10.1016/j.clim.2018.04.004 20. KONSTA OD, THABET Y, LE DANTEC C *et al*.:
- The contribution of epigenetics in Sjögren's syndrome. *Front Genet* 2014; 5: 71. https://doi.org/10.3389/fgene.2014.00071
- 21. TERUEL M, BARTUREN G, MARTINEZ-BUENO M *et al.*: Integrative epigenomics in Sjögren's syndrome reveals novel pathways and a strong interaction between the HLA, autoantibodies and the interferon signature. *Sci Rep* 2021; 11(1): 23292. https://doi.org/10.1038/s41598-021-01324-0
- ALTOROK N, COIT P, HUGHES T et al.: Genome-wide DNA methylation patterns in naive CD4<sup>+</sup> T cells from patients with primary Sjögren's syndrome. Arthritis Rheumatol 2014; 66(3): 731-9. https://doi.org/10.1002/art.38264
- IMGENBERG-KREUZ J, SANDLING JK, ALM-LÖF JC et al.: Genome-wide DNA methylation analysis in multiple tissues in primary Sjögren's syndrome reveals regulatory effects at interferon-induced genes. Ann Rheum Dis 2016; 75(11): 2029-36. https:// doi.org/10.1136/annrheumdis-2015-208659
- 24. CARNERO-MONTORO E, ALARCÓN-RIQUELME ME: Epigenome-wide association studies for systemic autoimmune diseases: The road behind and the road ahead. *Clin Immunol* 2018; 196: 21-33. https://doi.org/10.1016/j.clim.2018.03.014
- 25. YIN H, ZHAO M, WU X et al.: Hypomethylation and overexpression of CD70 (TNFSF7) in CD4+ T cells of patients with primary Sjögren's syndrome. J Dermatol Sci 2010; 59(3): 198-203. https://
- doi.org/10.1016/j.jdermsci.2010.06.011
   26. BROWNELL JE, ZHOU J, RANALLI T et al.: Tetrahymena histone acetyltransferase A: a homolog to yeast Gcn5p linking histone

- acetylation to gene activation. Cell 1996; 84(6): 843-51. https:// doi.org/10.1016/s0092-8674(00)81063-6
- 27. HUBER LC, STANCZYK J, JÜNGEL A, GAY S: Epigenetics in inflammatory rheumatic diseases. Arthritis Rheum 2007; 56(11): 3523-31. https://doi.org/10.1002/art.22948
- 28. SMOLLE M, WORKMAN JL: Transcriptionassociated histone modifications and cryptic transcription. Biochim Biophys Acta 2013; 1829(1): 84-97. https:// doi.org/10.1016/j.bbagrm.2012.08.008
- 29. CHOUDHARY C, WEINERT BT, NISHIDA Y, VERDIN E, MANN M: The growing landscape of lysine acetylation links metabolism and cell signalling. Nat Rev Mol Cell Biol 2014; 15(8): 536-50. https://doi.org/10.1038/nrm3841
- 30. DIEKER J, MULLER S: Epigenetic histone code and autoimmunity. Clin Rev Allergy Immunol 2010; 39(1): 78-84. https://doi.org/10.1007/s12016-009-8173-7
- 31. KONDO Y, SHEN L, CHENG AS et al.: Gene silencing in cancer by histone H3 lysine 27 trimethylation independent of promoter DNA methylation. Nat Genet 2008; 40(6): 741-50. https://doi.org/10.1038/ng.159
- 32. HOMBACH S, KRETZ M: Non-coding RNAs: Classification, Biology and Functioning. Adv Exp Med Biol 2016; 937: 3-17. https:// doi.org/10.1007/978-3-319-42059-2\_1
- 33. WEI JW, HUANG K, YANG C, KANG CS: Noncoding RNAs as regulators in epigenetics (Review). Oncol Rep 2017; 37(1): 3-9. https://doi.org/10.3892/or.2016.5236
- 34. SALIMINEJAD K, KHORRAM KHORSHID HR, SOLEYMANI FARD S, GHAFFARI SH: An overview of microRNAs: Biology, functions, therapeutics, and analysis methods. J Cell Physiol 2019; 234(5): 5451-65. https://doi.org/10.1002/jcp.27486
- 35. KABEKKODU SP, SHUKLA V, VARGHESE VK, J DS, CHAKRABARTY S, SATYAMOOR-THY K: Clustered miRNAs and their role in biological functions and diseases. Biol Rev Camb Philos Soc 2018; 93(4): 1955-86. https://doi.org/10.1111/brv.12428
- 36. MORENO-MOYA JM, VILELLA F, SIMÓN C: MicroRNA: key gene expression regulators. Fertil Steril 2014; 101(6): 1516-23. https:// doi.org/10.1016/j.fertnstert.2013.10.042
- 37. CHEN JQ, PAPP G, SZODORAY P, ZEHER M: The role of microRNAs in the pathogenesis of autoimmune diseases. Autoimmun Rev 2016; 15(12): 1171-80. https://doi.org/10.1016/j.autrev.2016.09.003
- 38. JARROUX J, MORILLON A, PINSKAYA M: History, Discovery, and Classification of IncRNAs. Adv Exp Med Biol 2017; 1008: 1-46. https:// doi.org/10.1007/978-981-10-5203-3\_1
- 39. CHOI SW, KIM HW, NAM JW: The small peptide world in long noncoding RNAs. Brief Bioinform 2019; 20(5): 1853-64. https://doi.org/10.1093/bib/bby055
- 40. BAK RO, MIKKELSEN JG: miRNA sponges: soaking up miRNAs for regulation of gene expression. Wiley Interdiscip Rev RNA 2014; 5(3): 317-33. https://doi.org/10.1002/wrna.1213
- 41. KARAGKOUNI D, KARAVANGELI A, PAR-ASKEVOPOULOU MD, HATZIGEORGIOU

- AG: Characterizing miRNA-lncRNA Interplay. Methods Mol Biol 2021; 2372: 243-62. https:// doi.org/10.1007/978-1-0716-1697-0\_21
- 42. FERRAGUT CARDOSO AP, BANERJEE M, NAIL AN, LYKOUDI A, STATES JC: miRNA dysregulation is an emerging modulator of genomic instability. Semin Cancer Biol 2021; 76: 120-31. https://
  - doi.org/10.1016/j.semcancer.2021.05.004
- 43. DE BENEDITTIS G. CICCACCI C. LATINI A, NOVELLI L, NOVELLI G, BORGIANI P: Emerging Role of microRNAs and Long Non-Coding RNAs in Sjögren's syndrome. Genes (Basel) 2021; 12(6). https://doi.org/10.3390/genes12060903
- 44. BALDINI C, CHATZIS LG, FULVIO G, LA ROCCA G, PONTARINI E, BOMBARDIERI M: Pathogenesis of Sjögren's disease: one year in review 2024. Clin Exp Rheumatol 2024; 42(12): 2336-43. https://
- doi.org/10.55563/clinexprheumatol/i8iszc 45. KAPSOGEORGOU EK, GOURZI VC, MAN-OUSSAKIS MN MOUTSOPOULOS HM TZI-OUFAS AG: Cellular microRNAs (miRNAs) and Sjögren's syndrome: candidate regulators of autoimmune response and autoantigen expression. J Autoimmun 2011; 37(2): 129-35.
- https://doi.org/10.1016/j.jaut.2011.05.003 46. IMGENBERG-KREUZ J, RASMUSSEN A, SIVILS K, NORDMARK G: Genetics and epigenetics in primary Sjögren's syndrome. Rheumatology (Oxford) 2021; 60(5): 2085-98. https://
- doi.org/10.1093/rheumatology/key330 47. PAULEY KM, STEWART CM, GAUNA AE et al.: Altered miR-146a expression in Sjögren's syndrome and its functional role in innate immunity. Eur J Immunol 2011; 41(7): 2029-39. https://doi.org/10.1002/eji.201040757
- 48. SHI H, ZHENG LY, ZHANG P, YU CQ: miR-146a and miR-155 expression in PBMCs from patients with Sjögren's syndrome. JOral pathol Med 2014; 43(10): 792-7. https://doi.org/10.1111/jop.12187
- 49. WANG Y, ZHANG G, ZHANG L, ZHAO M, HUANG H: Decreased microRNA-181a and -16 expression levels in the labial salivary glands of Sjögren syndrome patients. Exp Ther Med 2018; 15(1): 426-32 https://doi.org/10.3892/etm.2017.5407
- 50. LOPES AP, HILLEN MR, CHOURI E et al.: Circulating small non-coding RNAs reflect IFN status and B cell hyperactivity in patients with primary Sjögren's syndrome. PLoS One 2018; 13(2): e0193157. https:// doi.org/10.1371/journal.pone.0193157
- 51. RUSCA N, MONTICELLI S: MiR-146a in Immunity and Disease. Mol Biol Int 2011; 2011: 437301. https://doi.org/10.4061/2011/437301
- 52. ALEVIZOS I, ALEXANDER S, TURNER RJ, ILLEI GG: MicroRNA expression profiles as biomarkers of minor salivary gland inflammation and dysfunction in Sjögren's syndrome. Arthritis Rheum 2011; 63(2): 535-44. https://doi.org/ 10.1002/art.30131
- 53. ALEVIZOS I, ILLEI GG: MicroRNAs in Sjögren's syndrome as a prototypic autoimmune disease. Autoimmun Rev 2010; 9(9):

- 618-21. https:// doi.org/10.1016/j.autrev.2010.05.009
- 54. BRUNO D, TOLUSSO B, LUGLI G et al.: B-Cell Activation Biomarkers in Salivary Glands Are Related to Lymphomagenesis in Primary Sjögren's Disease: A Pilot Monocentric Exploratory Study. Int J Mol Sci 2024; 25(6). https://doi.org/10.3390/ijms25063259
- 55. WU H, CHEN Y, ZHU H, ZHAO M, LU Q: The Pathogenic Role of Dysregulated Epigenetic Modifications in Autoimmune Diseases. Front Immunol 2019; 10: 2305. https://doi.org/10.3389/fimmu.2019.02305
- 56. AGARWAL G, KUDAPA H, RAMALINGAM A et al.: Epigenetics and epigenomics: underlying mechanisms, relevance, and implications in crop improvement. Funct Integr Genomics 2020; 20(6): 739-61. https://doi.org/10.1007/s10142-020-00756-7
- 57. SHEN Y, YU X, WANG Q et al.: Association between primary Sjögren's syndrome and gut microbiota disruption: a systematic review and meta-analysis. Clin Rheumatol 2024; 43(2): 603-19. https:// doi.org/10.1007/s10067-023-06754-x
- 58. IGOE A, SCOFIELD RH: Autoimmunity and infection in Sjögren's syndrome. Curr Opin Rheumatol 2013; 25(4): 480-7. https:// doi.org/10.1097/BOR.0b013e32836200d2
- 59. BJÖRK A, MOFORS J, WAHREN-HERLENIUS M: Environmental factors in the pathogenesis of primary Sjögren's syndrome. J Inter Med 2020; 287(5): 475-92. https://doi.org/10.1111/joim.13032
- 60. TRIANTAFYLLOPOULOU A, TAPINOS N, MOUTSOPOULOS HM: Evidence for coxsackievirus infection in primary Sjögren's syndrome. Arthritis Rheum 2004; 50(9): 2897-902. https://doi.org/10.1002/art.20463
- 61. STATHOPOULOU EA, ROUTSIAS JG, STEA EA, MOUTSOPOULOS HM, TZIOUFAS AG: Cross-reaction between antibodies to the major epitope of Ro60 kD autoantigen and a homologous peptide of Coxsackie virus 2B protein. Clin Exp Immunol 2005; 141(1): 148-54. https://
- $\dot{doi.org/10.1111/j.1365-2249.2005.02812.x}$ 62. TUNG CH, LI CY, CHEN YC, CHEN YC: Association between nucleos(t)ide analogue therapy for hepatitis B and Sjögren's syndrome: 15-year analysis of the national database of Taiwan. J Viral Hepat 2021; 28(5): 809-16. https://doi.org/10.1111/jvh.13481
- 63. GALEOTTI C, BAYRY J: Autoimmune and inflammatory diseases following COVID-19. Nat Rev Rheumatol 2020; 16(8): 413-4. https://doi.org/10.1038/s41584-020-0448-7
- 64. HUDSON E, YANG L, CHU EK et al.: Evidence that autoantibody production may be driven by acute Epstein-Barr virus infection in Sjögren's disease. Ann Rheum Dis 2024 Oct. 29.
- https://doi.org/10.1136/ard-2024-226226 65. KABEERDOSS J, DEVARAJALU P, SAND-HYA P: DNA methylation profiling of labial salivary gland tissues revealed hypomethylation of B-cell-related genes in primary Sjögren's syndrome. Immunol Res 2024; 72(3): 450-9.
- https://doi.org/10.1007/s12026-024-09453-0 66. WANG Q, CHE N, LU C et al.: Correlation

- of peripheral CD4+GranzB+CTLs with disease severity in patients with primary Sjögren's syndrome. *Arthritis Res Ther* 2021; 23(1): 257. https://doi.org/10.1186/s13075-021-02632-6
- 67. DU Y, LI J, WU J, ZENG F, HE C: Exploration of the pathogenesis of Sjögren's syndrome via DNA methylation and transcriptome analyses. *Clin Rheumatol* 2022; 41(9): 2765-77. https://doi.org/10.1007/s10067-022-06200-4
- 68. BENCHABANE S, SLIMANI-KADDOURI A, ACHELI D, BENDIMERAD-IRATENE T, MESBAH R, TOUIL-BOUKOFFA C: Association between Increased Bcl-2, Fas and FasL Levels and Inflammation Extent in Labial Salivary Glands During Primary Sjögren's syndrome. Endocr Metab Immune Disord Drug Targets 2022; 22(3): 328-38. https:// doi.org/10.2174/1871530321666210809155147
- 69. MANTOVANI CARDOSO E, HUNDAL J, FETERMAN D, MAGALDI J: Concomitant new diagnosis of systemic lupus erythematosus and COVID-19 with possible antiphospholipid syndrome. Just a coincidence? A case report and review of intertwining pathophysiology. Clin Rheumatol 2020; 39(9): 2811-5.
  - https://doi.org/10.1007/s10067-020-05310-1
- CROIA C, ASTORRI E, MURRAY-BROWN W et al.: Implication of Epstein-Barr virus infection in disease-specific autoreactive B cell activation in ectopic lymphoid structures of Sjögren's syndrome. Arthritis Rheumatol 2014; 66(9): 2545-57. https://doi.org/10.1002/art.38726
- 71. GALLO A, JANG SI, ONG HL *et al.*: Targeting the Ca(2+) Sensor STIM1 by Exosomal Transfer of Ebv-miR-BART13-3p is Associated with Sjögren's syndrome. *EBioMedicine* 2016; 10: 216-26.
- https://doi.org/10.1016/j.ebiom.2016.06.041 72. GHOSH ROY S, ROBERTSON ES, SAHA A: Epigenetic Impact on EBV Associated B-Cell Lymphomagenesis. *Biomolecules* 2016; 6(4).
- https://doi.org/10.3390/biom6040046
- PAWLOTSKI JM, BEN YAHIA M, ANDRE C et al.: Immunological disorders in C virus chronic active hepatitis: a prospective case-control study. Hepatology 1994; 19(4): 841-8. https://doi.org/10.1016/0270-9139(94)90281-x
- 74. MARIETTE X, ZERBIB M, JACCARD A, SCHENMETZLER C, DANON F, CLAUVEL JP: Hepatitis C virus and Sjögren's syndrome.
- Arthritis Rheum 1993; 36(2): 280-1. https://doi.org/10.1002/art.1780360225 75. HADDAD J, DENY P, MUNZ-GOTHEIL C et al.: Lymphocytic sialadenitis of Sjögren's
- syndrome associated with chronic hepatitis C virus liver disease. *Lancet* 1992; 339(8789): 321-3. https://
- doi.org/10.1016/0140-6736(92)91645-o 76. PIRISI M, SCOTT C, FABRIS C *et al.*: Mild
- PIRISI M, SCOTT C, FABRIS C et al.: Mild sialoadenitis: a common finding in patients with hepatitis C virus infection. Scand J Gastroenterol 1994; 29(10): 940-2. https:// doi.org/10.3109/00365529409094867
- RAMOS-CASALS M, FONT J: Extrahepatic manifestations in patients with chronic hepatitis C virus infection. Curr Opin Rheuma-

- tol 2005; 17(4): 447-55. https://doi.org/10.1097/01.bor.0000166386.62851.49
- SHIBOSKI CH, SHIBOSKI SC, SEROR R et al.: 2016 American College of Rheumatology/European League Against Rheumatism Classification Criteria for Primary Sjögren's Syndrome: A Consensus and Data-Driven Methodology Involving Three International Patient Cohorts. Ann Rheum Dis 2017; 69(1): 35-45. https://doi.org/10.1136/annrheumdis-2016-210571
- 79. POLYAK SJ, CRISPE IN, BAUMERT TF: Liver Abnormalities after Elimination of HCV Infection: Persistent Epigenetic and Immunological Perturbations Post-Cure. *Pathogens* 2021; 10(1).
- https://doi.org/10.3390/pathogens10010044 80. NAWAZ R, ZAHID S, IDREES M *et al.*: HCV-induced regulatory alterations of IL-1β, IL-6, TNF-α, and IFN-γ operative, leading liver en-route to non-alcoholic steatohepatitis. *Inflamm Res* 2017; 66(6): 477-86. https://doi.org/10.1007/s00011-017-1029-3
- UMEKITA K, OKAYAMA A: HTLV-1 Infection and Rheumatic Diseases. Front Microbiol 2020; 11: 152. https://doi.org/10.3389/fmicb.2020.00152
- MATSUOKA M, MESNARD JM: HTLV-1 bZIP factor: the key viral gene for pathogenesis. *Retrovirology* 2020; 17(1): 2. https://doi.org/10.1186/s12977-020-0511-0
- NAKAMURA H, SHIMIZU T, TAKAGI Y et al.: Reevaluation for clinical manifestations of HTLV-I-seropositive patients with Sjögren's syndrome. BMC Musculoskelet Disord 2015; 16: 335.
- https://doi.org/10.1186/s12891-015-0773-1 84. GREEN JE, HINRICHS SH, VOGEL J, JAY G: Exocrinopathy resembling Sjögren's syndrome in HTLV-1 tax transgenic mice. *Nature* 1989; 341(6237): 72-4. https://doi.org/10.1038/341072a0
- 85. MARIETTE X, AGBALIKA F, ZUCKER-FRANKLIN D *et al.*: Detection of the tax gene of HTLV-I in labial salivary glands from patients with Sjögren's syndrome and other diseases of the oral cavity. *Clin Exp Rheumatol* 2000; 18(3): 341-7.
- 86. TERADA K, KATAMINE S, EGUCHI K et al.: Prevalence of serum and salivary antibodies to HTLV-1 in Sjögren's syndrome. Lancet 1994; 344(8930): 1116-9. https:// doi.org/10.1016/s0140-6736(94)90630-0
- 87. NAKAMURA H, TAKAGI Y, KAWAKAMI A, IDA H, NAKAMURA T, EGUCHI K: HTLV-I infection results in resistance toward salivary gland destruction of Sjögren's syndrome. Clin Exp Rheumatol 2008; 26(4): 653-5.
- 88. VALE DAD, CASSEB J, DE OLIVEIRA ACP, BUSSOLOTI FILHO I, DE SOUSA SCOM, ORTEGA KL: Prevalence of Sjögren's syndrome in Brazilian patients infected with human T-cell lymphotropic virus. *J Oral* Pathol Med 2017; 46(7): 543-8. https://doi.org/10.1111/jop.12530
- 89. NAKAMURA H, SHIMIZU T, KAWAKAMU A: Role of Viral Infections in the Pathogenesis of Sjögren's Syndrome: Different Characteristics of Epstein-Barr Virus and HTLV-1. *J Clin Med* 2020; 9(5). https://doi.org/10.3390/jcm9051459

- NAKAMURA H, KAWAKAMI A, HAYASHI T et al.: Low prevalence of ectopic germinal centre formation in patients with HTLV-I-associated Sjögren's syndrome. Rheumatology (Oxford) 2009; 48(7): 854-5. https://doi.org/10.1093/rheumatology/kep072
- 91. EMERY VC: Investigation of CMV disease in immunocompromised patients. *J Clin Pathol* 2001; 54(2): 84-8. https://doi.org/10.1136/jcp.54.2.84
- 92. LEE Y, RISKEDAL E, KALLEBERG KT *et al.*: EWAS of post-COVID-19 patients shows
- methylation differences in the immuneresponse associated gene, IFI44L, three months after COVID-19 infection. *Sci Rep* 2022; 12(1): 11478.
- https://doi.org/10.1038/s41598-022-15467-1
- BALADA E, ORDI-ROS J, VILARDELL-TAR-RÉS M: Molecular mechanisms mediated by human endogenous retroviruses (HERVs) in autoimmunity. Rev Med Virol 2009; 19(5): 273-86. https://doi.org/10.1002/rmv.622
- 94. DANTEC LC, CHARRAS A, BROOKS WH, RENAUDINEAU Y: Similarities and Differences of Epigenetic Mechanisms in Lupus and Sjogren's syndrome. *Lupus Open Ac*cess 2015; 1: e101. https:// doi.org/10.35248/2684-1630.16.1.e101
- 95. MAVRAGANI CP, SAGALOVSKIY I, GUO Q et al.: Expression of Long Interspersed Nuclear Element 1 Retroelements and Induction of Type I Interferon in Patients with Systemic Autoimmune Disease. Arthritis Rheumatol 2016; 68(11): 2686-96. https://doi.org/10.1002/art.39795
- 96. MAVRAGANI CP, NEZOS A, SAGALOVSKIY I, SESHAN S, KIROU KA, CROW MK: Defective regulation of L1 endogenous retroelements in primary Sjögren's syndrome and systemic lupus erythematosus: Role of methylating enzymes. *J Autoimmun* 2018; 88: 75-82.
  - https://doi.org/10.1016/j.jaut.2017.10.004
- 97. THABET Y, LE DANTEC C, GHEDIRA I *et al.*: Epigenetic dysregulation in salivary glands from patients with primary Sjögren's syndrome may be ascribed to infiltrating B cells. *J Autoimmun* 2013; 41: 175-81. https://doi.org/10.1016/j.jaut.2013.02.002
- 98. LUQ, RENAUDINEAU Y, CHAS *et al.*: Epigenetics in autoimmune disorders: highlights of the 10th Sjögren's syndrome symposium. *Autoimmun Rev* 2010; 9(9): 627-30. https://doi.org/10.1016/j.autrev.2010.05.011
- 99. GARAUD S, LE DANTEC C, JOUSSE-JOULIN S *et al.*: IL-6 modulates CD5 expression in B cells from patients with lupus by regulating DNA methylation. *J Immunol* 2009; 182(9): 5623-32. https://doi.org/10.4049/jimmunol.0802412
- 100. RENAUDINEAU Y, GARAUD S, LE DANTEC C, ALONSO-RAMIREZ R, DARIDON C, YOUINOU P: Autoreactive B cells and epigenetics. Clin Rev Allergy Immunol 2010; 39(1): 85-94.
- https://doi.org/10.1007/s12016-009-8174-6
  101. CUI D, AN R, LI L, JIANG L, JIANG C, JIN J:
  Causal association between Helicobacter
  pylori infection and Sjogren's syndrome:
  a bidirectional Mendelian randomization
  analysis. BMC Infect Dis 2024; 24(1): 782.
  https://doi.org/10.1186/s12879-024-09678-2

- 102. CHEN Q, ZHOU X, TAN W, ZHANG M: Association between Helicobacter pylori infection and Sjögren syndrome: A meta-analysis. *Medicine* (Baltimore) 2018; 97(49): e13528. https://doi.org/10.1097/MD.00000000000013528
- 103. FLOCH P, CAPDEVIELLE C, STAEDEL C et al.: Deregulation of MicroRNAs in Gastric Lymphomagenesis Induced in the d3Tx Mouse Model of Helicobacter pylori Infection. Front Cell Infect Microbiol 2017; 7:
- https://doi.org/10.3389/fcimb.2017.00185 104. DOW CT, CHAN ED: What is the evidence that mycobacteria are associated with the pathogenesis of Sjögren's syndrome? *J Transl Autoimmun* 2021; 4: 100085.
- https://doi.org/10.1016/j.jtauto.2021.100085 105. RADIĆ M, KOLAK E, ĐOGAŠ H *et al.*: Vitamin D and Sjögren's Disease: Revealing the Connections-A Systematic Review and Meta-Analysis. *Nutrients* 2023; 15(3). https://doi.org/10.3390/nu15030497
- 106. LI L, CHEN J, JIANG Y: The association between vitamin D level and Sjögren's syndrome: A meta-analysis. *Int J Rheum Dis* 2019; 22(3): 532-3. https://doi.org/10.1111/1756-185X.13474
- 107. ILLESCAS-MONTES R, MELGUIZO-RODRÍ-GUEZ L, RUIZ C, COSTELA-RUIZ VJ: Vitamin D and autoimmune diseases. *Life Sci* 2019; 233: 116744.
- https://doi.org/10.1016/j.lfs.2019.116744 108. BARTLEY J: Vitamin D: emerging roles in infection and immunity. Expert Rev Anti Infect Ther 2010; 8(12): 1359-69. https://doi.org/10.1586/eri.10.102
- 109. KENNEL KA, DRAKE MT, HURLEY DL: Vitamin D deficiency in adults: when to test and how to treat. *Mayo Clin Proc* 2010; 85(8): 752-7; quiz 7-8. https://doi.org/10.4065/mcp.2010.0138
- 110. HENDI NN, NEMER G: Epigenetic regulation of vitamin D deficiency. *Epigenomics* 2023; 15(12): 653-5. https://doi.org/10.2217/epi-2023-0246
- 111. COSTENBADER KH, COOK NR, LEE IM et al.: Vitamin D and Marine n-3 Fatty Acids for Autoimmune Disease Prevention: Outcomes Two Years After Completion of a Double-Blind, Placebo-Controlled Trial. Arthritis Rheumatol 2024; 76(6): 973-83. https://doi.org/10.1002/art.42811
- 112. STRICKLAND FM, HEWAGAMA A, LU Q *et al.*: Environmental exposure, estrogen and two X chromosomes are required for disease development in an epigenetic model of lupus. *J Autoimmun* 2012; 38(2-3): J135-43. https://doi.org/10.1016/j.jaut.2011.11.001
- 113. WU Z, LI X, QIN H, ZHU X, XU J, SHI W: Ultraviolet B enhances DNA hypomethylation of CD4+ T cells in systemic lupus erythematosus via inhibiting DNMT1 catalytic activity. *J Dermatol Sci* 2013; 71(3): 167-73. https://
- doi.org/10.1016/j.jdermsci.2013.04.022
  114. MAZUR A, FRĄCZEK P, TABARKIEWICZ
  J: Vitamin D as a Nutri-Epigenetic Factor in Autoimmunity-A Review of Current Research and Reports on Vitamin D Deficiency in Autoimmune Diseases. *Nutrients* 2022; 14(20).

- https://doi.org/10.3390/nu14204286
- 115. DABRAVOLSKI SA, CHUROV AV, STARO-DUBTSEVA IA et al.: Vitamin D in Primary Sjögren's Syndrome (pSS) and the Identification of Novel Single-Nucleotide Polymorphisms Involved in the Development of pSS-Associated Diseases. *Diagnostics* 2024; 14(18). https:// doi.org/10.3390/diagnostics14182035
- 116. TROMBETTA AC, SMITH V, GOTELLI E et al.: Vitamin D deficiency and clinical correlations in systemic sclerosis patients: A retrospective analysis for possible future developments. PLoS One 2017; 12(6): e0179062. https://doi.org/10.1371/journal.pone.0179062
- 117. VERNIA F, VALVANO M, LONGO S, CESARO N, VISCIDO A, LATELLA G: Vitamin D in Inflammatory Bowel Diseases. Mechanisms of Action and Therapeutic Implications. *Nutrients* 2022; 14(2). https://doi.org/10.3390/nu14020269
- 118. JIN L, DAI M, LI C, WANG J, WU B: Risk factors for primary Sjögren's Syndrome: a systematic review and meta-analysis. *Clin Rheumatol* 2023; 42(2): 327-38. https://doi.org/10.1007/s10067-022-06474-8
- 119. MOFORS J, BJÖRK A, RICHARDSDOTTER ANDERSSON E *et al.*: Cigarette smoking patterns preceding primary Sjögren's syndrome. *RMD Open* 2020; 6(3). https://doi.org/10.1136/rmdopen-2020-001402
- 120. BEN-ELI H, AFRAMIAN DJ, BEN-CHETRIT E et al.: Shared Medical and Environmental Risk Factors in Dry Eye Syndrome, Sjögren's Syndrome, and B-Cell Non-Hodgkin Lymphoma: A Case-Control Study. J Immunol Res 2019; 2019: 9060842. https://doi.org/10.1155/2019/9060842
- 121. OLSSON P, TURESSON C, MANDL T, JACOB-SSON L, THEANDER E: Cigarette smoking and the risk of primary Sjögren's syndrome: a nested case control study. *Arthritis Res Ther* 2017; 19(1): 50. https://doi.org/10.1186/s13075-017-1255-7
- 122. PRIORI R, MEDDA E, CONTI F et al.: Risk factors for Sjögren's syndrome: a case-control study. Clin Exp Rheumatol 2007; 25(3): 378-84
- 123. ANDERSEN A, REIMER R, DAWES K *et al.*:
  DNA methylation differentiates smoking from vaping and non-combustible tobacco use. *Epigenetics* 2022; 17(2): 178-90. https://doi.org/10.1080/15592294.2021.1890875
- 124. RICHMOND RC, SILLERO-REJON C, KHOU-JA JN et al.: Investigating the DNA methylation profile of e-cigarette use. Clin Epigenetics 2021; 13(1): 183. https:// doi.org/10.1186/s13148-021-01174-7
- 125. ZEILINGER S, KÜHNEL B, KLOPP N et al.: Tobacco smoking leads to extensive genome-wide changes in DNA methylation. PLoS One 2013; 8(5): e63812. https://doi.org/10.1371/journal.pone.0063812
- 126. PERRICONE C, VERSINI M, BEN-AMI D *et al.*: Smoke and autoimmunity: The fire behind the disease. *Autoimmun Rev* 2016; 15(4): 354-74. https://doi.org/10.1016/j.autrev.2016.01.001
- 127. HURTADO C, ACEVEDO SÁENZ LY, VÁSQUEZ TRESPALACIOS EM et al.: DNA

- methylation changes on immune cells in Systemic Lupus Erythematosus. *Autoimmunity* 2020; 53(3): 114-21. https://doi.org/10.1080/08916934.2020.1722108
- 128. TSAPROUNI LG, YANG TP, BELL J et al.: Cigarette smoking reduces DNA methylation levels at multiple genomic loci but the effect is partially reversible upon cessation. Epigenetics 2014; 9(10): 1382-96. https://doi.org/10.4161/15592294.2014.969637
- 129. BARBHAIYA M, COSTENBADER KH: Environmental exposures and the development of systemic lupus erythematosus. *Curr Opin Rheumatol* 2016; 28(5): 497-505. https://doi.org/10.1097/BOR.0000000000000318
- 130. SOMERS EC, RICHARDSON BC: Environmental exposures, epigenetic changes and the risk of lupus. *Lupus* 2014; 23(6): 568-76. https://doi.org/10.1177/0961203313499419
- 131. MAHNKE AH, MIRANDA RC, HOMANICS GE: Epigenetic mediators and consequences of excessive alcohol consumption. *Alcohol* 2017; 60: 1-6. https://doi.org/10.1016/j.alcohol.2017.02.357
- 132. CURTIS BJ, ZAHS A, KOVACS EJ: Epigenetic targets for reversing immune defects caused by alcohol exposure. *Alcohol Res* 2013; 35(1): 97-113. https://doi.org/10.35946/arcr.v35.1.11
- 133. TERRACINA S, TARANI L, CECCANTI M et al.: The Impact of Oxidative Stress on the Epigenetics of Fetal Alcohol Spectrum Disorders. Antioxidants (Basel) 2024; 13(4). https://doi.org/10.3390/antiox13040410
- 134. TERRACINA S, FERRAGUTI G, TARANI L et al.: Transgenerational Abnormalities Induced by Paternal Preconceptual Alcohol Drinking: Findings from Humans and Animal Models. Curr Neuropharmacol 2022; 20(6): 1158-73. https:// doi.org/10.2174/1570159X19666211101111430
- 135. WIRLEITNER B, SCHROECKSNADEL K, WINKLER C, SCHENNACH H, FUCHS D: Resveratrol suppresses interferon-gamma-induced biochemical pathways in human peripheral blood mononuclear cells in vitro. Immunol Lett 2005; 100(2): 159-63. https://doi.org/10.1016/j.imlet.2005.03.008
- 136. PETRELLA C, CARITO V, CARERE C et al.:
  Oxidative stress inhibition by resveratrol in alcohol-dependent mice. Nutrition 2020; 79-80: 110783.
  - https://doi.org/10.1016/j.nut.2020.110783
- 137. MATHERS JC, STRATHDEE G, RELTON CL: Induction of epigenetic alterations by dietary and other environmental factors. *Adv Genet* 2010; 71: 3-39. https://doi.org/10.1016/B978-0-12-380864-6.00001-8
- 138. KIYOHARA C, WASHIO M, HORIUCHI T et al.: Modifying effect of N-acetyltransferase 2 genotype on the association between systemic lupus erythematosus and consumption of alcohol and caffeine-rich beverages. Arthritis Care Res (Hoboken) 2014; 66(7): 1048-56. https://doi.org/10.1002/acr.22282
- 139. CORNACCHIA E, GOLBUS J, MAYBAUM J, STRAHLER J, HANASH S, RICHARDSON B: Hydralazine and procainamide inhibit T cell DNA methylation and induce autoreactivity. *J Immunol* 1988; 140(7): 2197-200. https://doi.org/10.4049/jimmunol.140.7.2197
- 140. RICHARDSON B: Primer: epigenetics of

- autoimmunity. *Nat Clin Pract Rheumatol* 2007; 3(9): 521-7. https://doi.org/10.1038/ncprheum0573
- nttps://doi.org/10.1038/ncprneum03/3
- 141. SCHEINBART LS, JOHNSON MA, GROSS LA, EDELSYEIN SR, RICHARDSON BC: Procainamide inhibits DNA methyltransferase in a human T cell line. *J Rheumatol* 1991; 18(4): 530-4.
- 142. DENG C, LU Q, ZHANG Z *et al.*: Hydralazine may induce autoimmunity by inhibiting extracellular signal-regulated kinase pathway signaling. *Arthritis Rheum* 2003; 48(3): 746-56. https://doi.org/10.1002/art.10833
- 143. YUNG R, POWERS D, JOHNSON K et al.: Mechanisms of drug-induced lupus. II. T cells overexpressing lymphocyte function-associated antigen 1 become autoreactive and cause a lupuslike disease in syngeneic mice. J Clin Invest 1996; 97(12): 2866-71. https://doi.org/10.1172/JCI118743
- 144. ALEGRÍA-TORRES JA, BACCARELLI A, BOLLATI V: Epigenetics and lifestyle. *Epigenomics* 2011; 3(3): 267-77.

- https://doi.org/10.2217/epi.11.22
- 145. GORELIK GJ, YARLAGADDA S, PATEL DR, RICHARDSON BC: Protein kinase Cδ oxidation contributes to ERK inactivation in lupus T cells. *Arthritis Rheum* 2012; 64(9): 2964-74. https://doi.org/10.1002/art.34503
- 146. AKHIL A, BANSAL R, ANUPAM K, TANDON A, BHATNAGAR A: Systemic lupus erythematosus: latest insight into etiopathogenesis. *Rheumatol Int* 2023; 43(8): 1381-93. https://doi.org/10.1007/s00296-023-05346-x
- 147. MAHMOUD AM, ALI MM: Methyl Donor Micronutrients that Modify DNA Methylation and Cancer Outcome. *Nutrients* 2019; 11(3). https://doi.org/10.3390/nu11030608
- 148. GILCREASE GW, PADOVAN D, HEFFLER E et al.: Is air pollution affecting the disease activity in patients with systemic lupus erythematosus? State of the art and a systematic literature review. Eur J Rheumatol 2020; 7(1): 31-4. https://
- doi.org/10.5152/eurjrheum.2019.19141 149. HORAI Y, SHIMIZU T, UMEDA M, NISHIHATA

- SY, NAKAMURA H, KAWAKAMI A: Current Views on Pathophysiology and Potential Therapeutic Targets in Sjögren's syndrome: A Review from the Perspective of Viral Infections, Toll-like Receptors, and Long-Noncoding RNAs. *J Clin Med* 2023; 12(18). https://doi.org/10.3390/jcm12185873
- 150. THORLACIUS GE, BJÖRK A, WAHREN-HERLENIUS M: Genetics and epigenetics of primary Sjögren syndrome: implications for future therapies. *Nat Rev Rheumatol* 2023; 19(5): 288-306.
- https://doi.org/10.1038/s41584-023-00932-6 151. BAE SC, LEE YH: Association between Vitamin D level and/or deficiency, and systemic lupus erythematosus: a meta-analysis. *Cell Mol Biol* (Noisy-le-grand) 2018; 64(1):
- https://doi.org/10.14715/cmb/2018.64.1.2 152. GORELIK G, RICHARDSON B: Key role of ERK pathway signaling in lupus. *Autoim-munity* 2010; 43(1): 17-22. https:// doi.org/10.3109/08916930903374832